

Versatility of the CoPoP platform for antigen discovery and tuberculosis vaccine testing

Andres Obregon-Henao¹, Yang Jiao², Kristina Tran¹, Wen Ling Hsu², Brennen Troyer¹, Marcela I Henao-Tamayo¹, Jonathan F Lovell²

¹Colorado State University, Fort Collins, Colorado, USA; ²University at Buffalo, Buffalo, New York, USA

Introduction: Tuberculosis (TB) continues to plague humankind. The Covid-19 pandemic erased years of progress in our fight against TB, highlighting the need to innovate in every facet of this disease: diagnosis, treatment, and vaccine development. Specifically, the century-old BCG —a live attenuated M.bovis strain— remains the only licensed vaccine, despite its limited protection against pulmonary TB. Recent studies show subunit proteins can be used as prime vaccines or to boost BCG protection. Herein, we used Cobalt Porphyrin Phospholipid (CoPoP) liposome platform developed by Lovell, for antigen discovery and TB vaccine testing using recombinant Mtb his-tagged proteins and/or synthetic peptides.

Methods: Synthetic his-tagged peptides and recombinant Mtb proteins were obtained commercially or in-house (Lovell lab and CSU's protein core). Peptide sequences were obtained from literature search or bioinformatic analysis of TB proteins via NetMHCII. Vaccines were generated by co-incubating his-tagged proteins/peptides + CoPoP in optimized conditions. Female mice were prime vaccinated IM with CoPoP or subQ BCG Pasteur, followed by CoPoP boosting 3 weeks later. Bacterial burden enumeration and histopathological analysis was performed in lungs and spleens 1 or 3 months after aerosol infection with Mtb H37Rv.

Results. More than 50 antigens (peptides/recombinant proteins) have been screened in 2 years. Several individual antigens induced protection (lower CFUs and pathology) at 1 month, potentially synergizing when multiplexed. Ongoing studies are evaluating long-term protection by multiplexed vaccines and/or as BCG boost.

Discussion. CoPoP represent a versatile vaccine platform for TB antigen discovery and vaccine testing. Versatility is reflected in multiple ways: ease to manufacture/store; compatibility with single/fusion/multiplexed antigens, in-house prepped or commercially available his-tagged proteins, and synthetic peptides from difficult to express Mtb proteins.

Funding Sources

NIH R61AI169199 to AOH and JFL

Conflicts of Interest None

